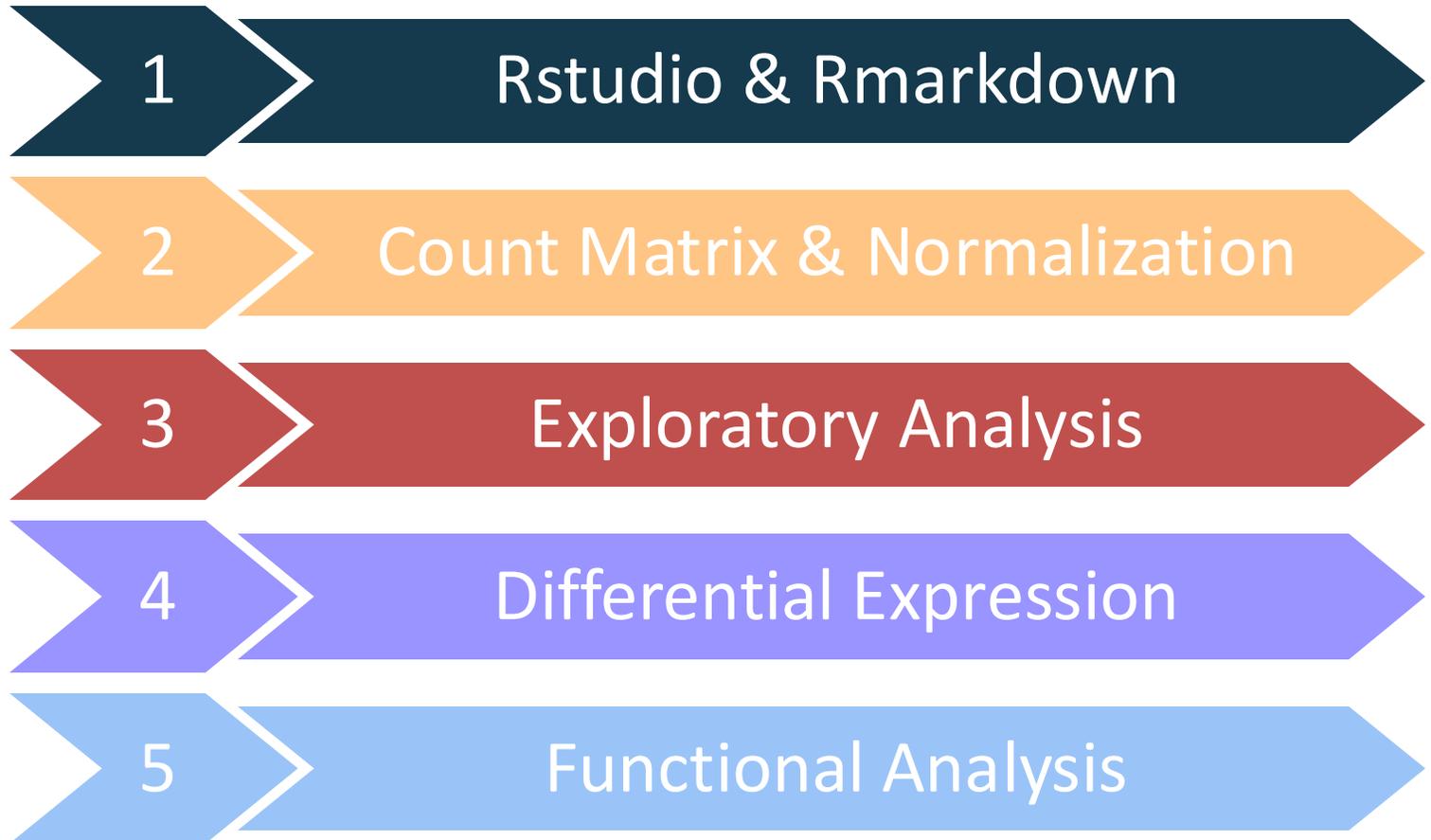


Functional Analysis

Center for Health Data Science



Overview

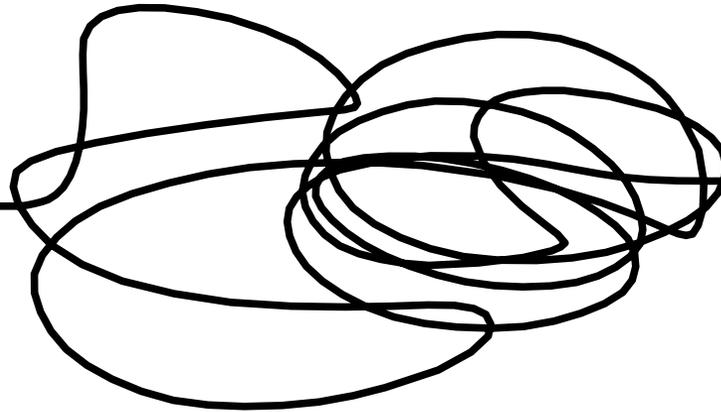


Functional analysis

- Differential Expression analysis
 - Resulted in **up**- and **down**-regulated genes in each comparison
- Define genes of interest:
 - Treatment vs. Control with $\text{Log}_2\text{FC} > 1$ and adjusted p-value < 0.05
- What do these genes do?
 - Do they share a **common function**? e.g. Immuno-processes
 - Part of the **same pathway**? e.g. Nucleotide Metabolism

Functional analysis

1. Differentially Expressed Genes



3. Publish!

2. Anything and everything biologically meaningful & interesting:

- Co-expression and interaction
- Gene set enrichment analysis (pathway, GO terms)
- Disease and drug databases
- ...

Annotation

DESeq2 results:

Gene_ID (Ensembl)	LFC	padj	...
<u>ENSG00000223972</u>	1.101	0.001	...
<u>ENSG00000278267</u>	-4.567	0.045	...

Annotation:

EBI (Ensembl)

HGNC

NCBI

UCSC

Gene_ID (Ensembl)	Gene_name	Entrez_ID	RefSeq_ID	Chr	Start	End	Feature
<u>ENSG00000223972</u>	DDX11L1	NA	NR_046018	1	11869	14409	Protein
<u>ENSG00000278267</u>	MIR6859-1	102466751	NR_106918	1	17369	17436	nc-RNA

+ MANE = Matched Annotation between NCBI and EBI

Annotation

Genome assembly updated **every few years**
Annotations updated **every few months**

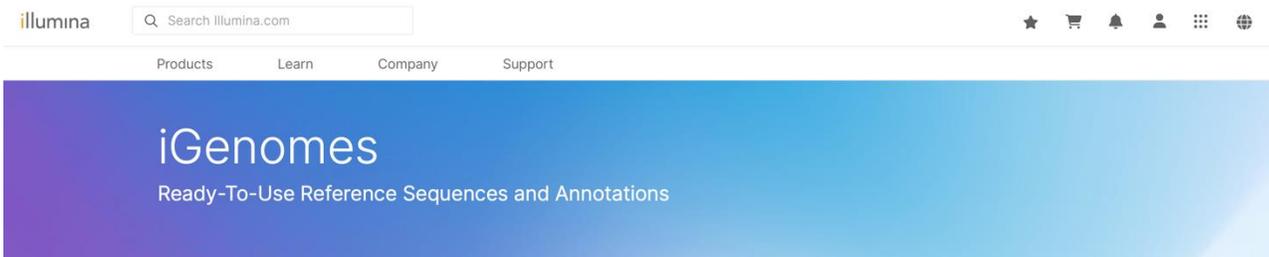
⚠ **Genome assembly versions MUST match!**

- If you map to **GRCh37**, use annotations for **GRCh37**
- Preferably also use the **same release** for all annotations

Ensembl database annotation releases for GRCh37 build:

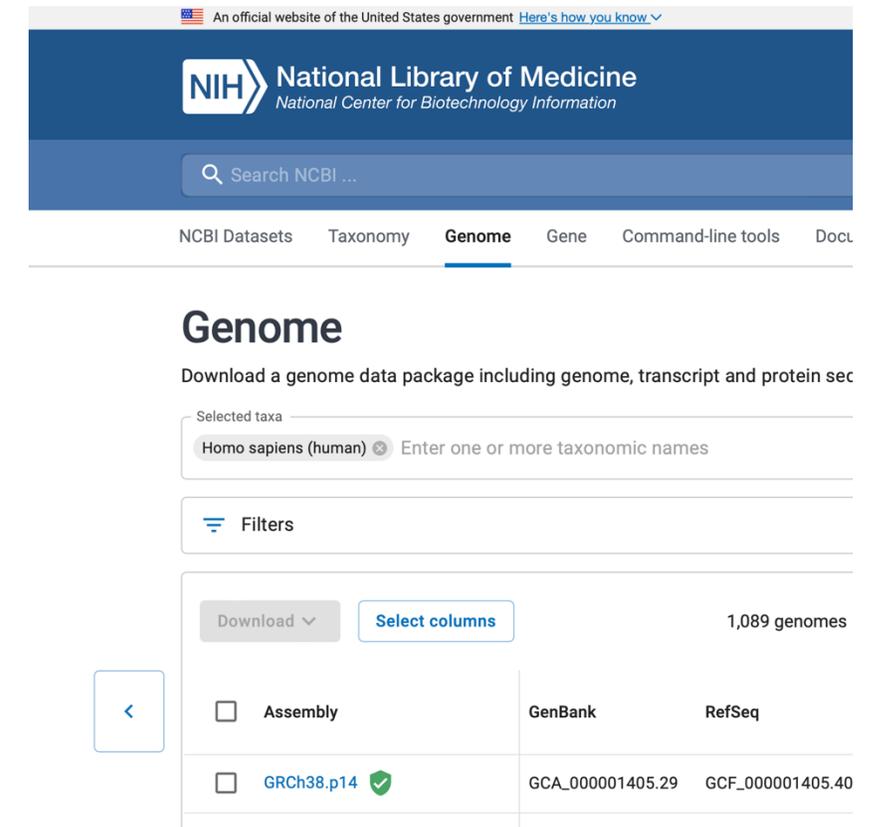
```
> listEnsemblArchives()
  name      date      url      version current_release
1 Ensembl GRCh37 Feb 2014 https://grch37.ensembl.org GRCh37
2 Ensembl 108 Oct 2022 https://oct2022.archive.ensembl.org 108 *
3 Ensembl 107 Jul 2022 https://jul2022.archive.ensembl.org 107
4 Ensembl 106 Apr 2022 https://apr2022.archive.ensembl.org 106
5 Ensembl 105 Dec 2021 https://dec2021.archive.ensembl.org 105
6 Ensembl 104 May 2021 https://may2021.archive.ensembl.org 104
7 Ensembl 103 Feb 2021 https://feb2021.archive.ensembl.org 103
8 Ensembl 102 Nov 2020 https://nov2020.archive.ensembl.org 102
9 Ensembl 101 Aug 2020 https://aug2020.archive.ensembl.org 101
10 Ensembl 100 Apr 2020 https://apr2020.archive.ensembl.org 100
11 Ensembl 99 Jan 2020 https://jan2020.archive.ensembl.org 99
12 Ensembl 98 Sep 2019 https://sep2019.archive.ensembl.org 98
13 Ensembl 97 Jul 2019 https://jul2019.archive.ensembl.org 97
14 Ensembl 96 Apr 2019 https://apr2019.archive.ensembl.org 96
15 Ensembl 95 Jan 2019 https://jan2019.archive.ensembl.org 95
16 Ensembl 94 Oct 2018 https://oct2018.archive.ensembl.org 94
17 Ensembl 93 Jul 2018 https://jul2018.archive.ensembl.org 93
18 Ensembl 92 Apr 2018 https://apr2018.archive.ensembl.org 92
19 Ensembl 91 Dec 2017 https://dec2017.archive.ensembl.org 91
20 Ensembl 80 May 2015 https://may2015.archive.ensembl.org 80
21 Ensembl 77 Oct 2014 https://oct2014.archive.ensembl.org 77
22 Ensembl 75 Feb 2014 https://feb2014.archive.ensembl.org 75
23 Ensembl 54 May 2009 https://may2009.archive.ensembl.org 54
```

Annotation



 A resource for downloading matching references + annotations

A more updated resource 



Annotation

Reference genome = gold standard

Genome assembly GRCh38.p14 reference



- **Chromosome level** assembly
- Contains **chromosomes + scaffolds** with funny names
- Some regions **contain gaps** (telomeres, repetitive regions, centromeres)

Newer version but less explored

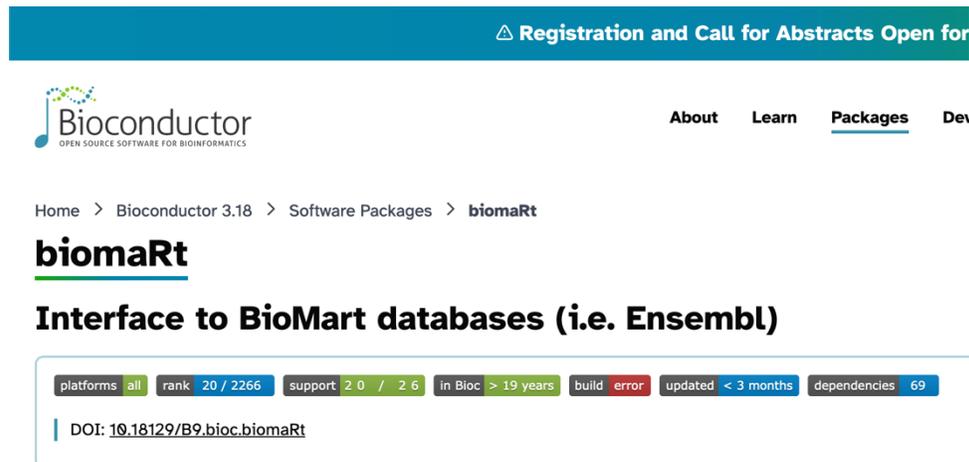
Genome assembly T2T-CHM13v2.0



- **Whole genome** assembly
- Contains **only chromosomes**
- **Complete sequence** from telomere to telomere

Annotation in R

- R packages **biomaRt** or **AnnotationDbi** can be used to convert between identifiers
- You *may* not be able to get the exact release version of a build, but you can likely get one close to it:
 - This could mean some IDs cannot be converted, it is only a few we usually don't care



Registration and Call for Abstracts Open for

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About Learn Packages Dev

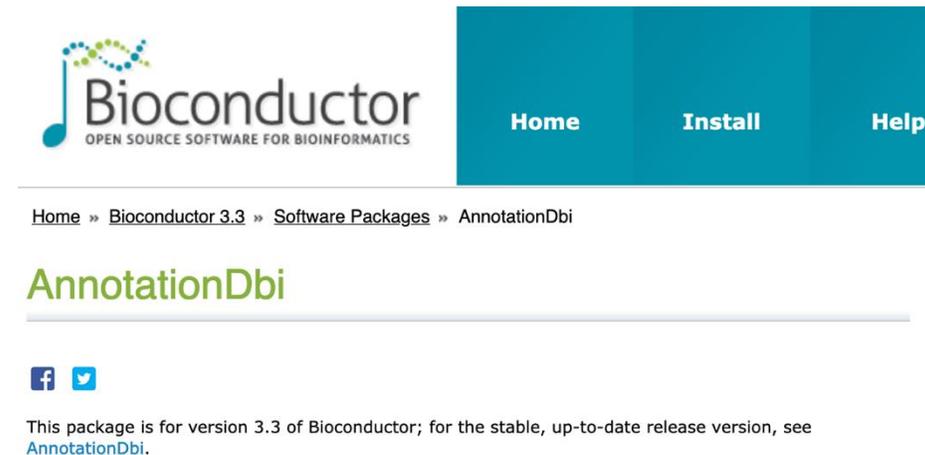
Home > Bioconductor 3.18 > Software Packages > **biomaRt**

biomaRt

Interface to BioMart databases (i.e. Ensembl)

platforms all rank 20 / 2266 support 2.0 / 2.6 in Bioc > 19 years build error updated < 3 months dependencies 69

DOI: [10.18129/B9.bioc.biomaRt](https://doi.org/10.18129/B9.bioc.biomaRt)



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Home Install Help

Home » Bioconductor 3.3 » Software Packages » AnnotationDbi

AnnotationDbi

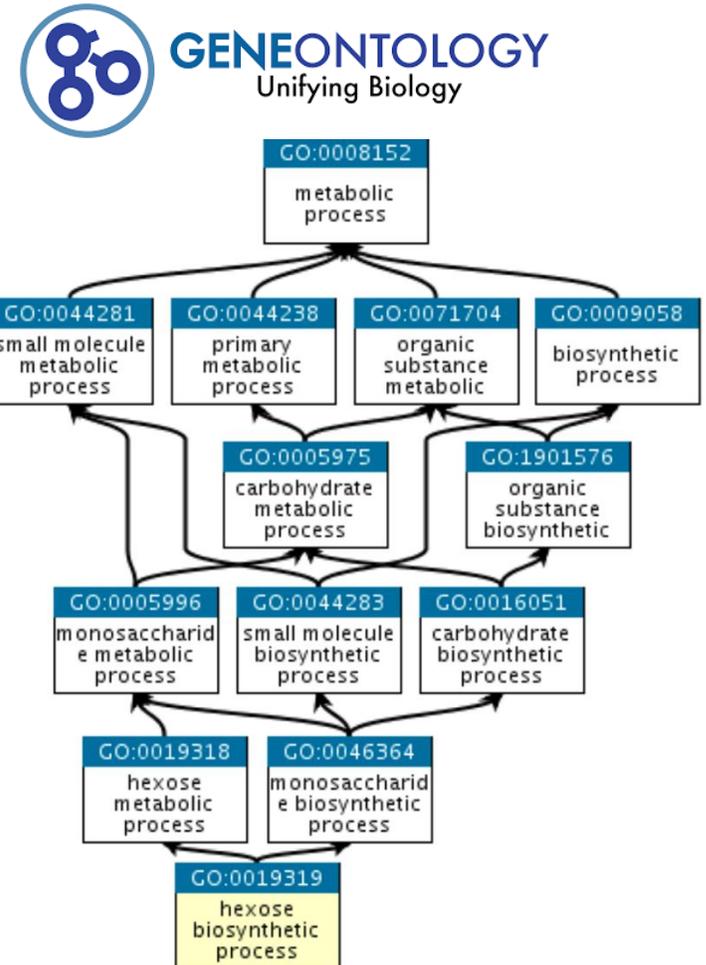
f t

This package is for version 3.3 of Bioconductor; for the stable, up-to-date release version, see [AnnotationDbi](#).

Functional analysis – GO terms

Gene Ontology (GO) Term

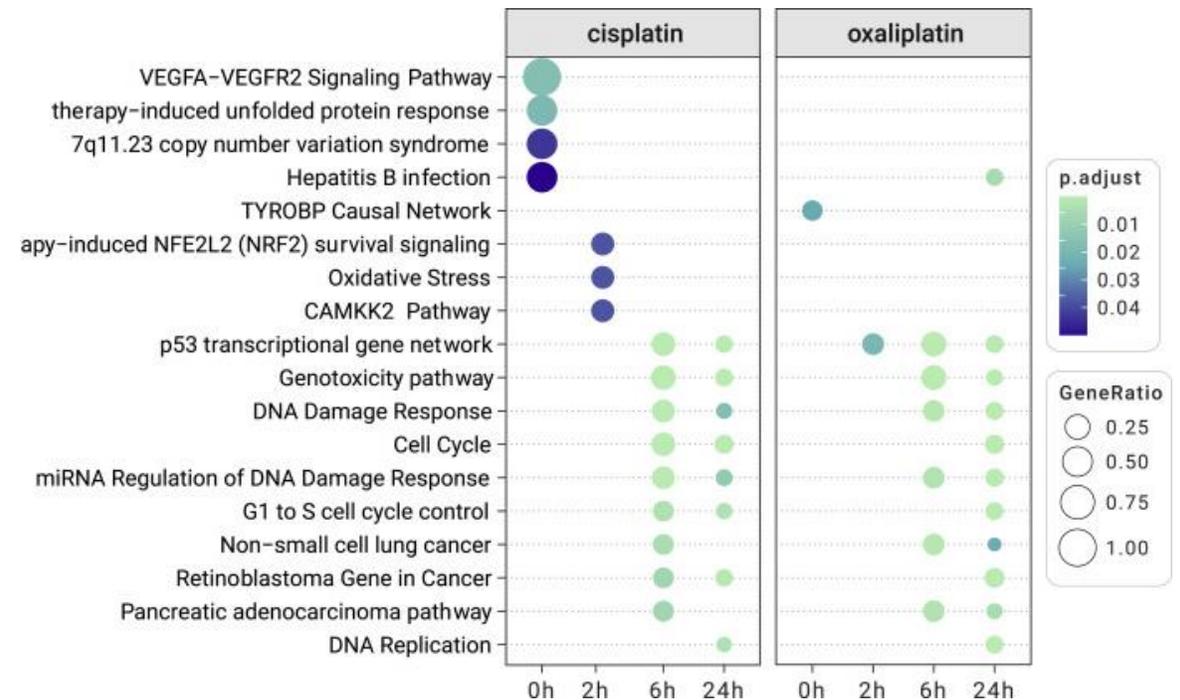
- Formal representation of a body of knowledge in the biological domain
- Genes are annotated to different types of knowledge
 - Biological processes: DNA repair, signal transduction, etc.
 - Molecular function: catalysis, transportation, etc.
 - Cellular component: ribosome, nucleus, etc.



Functional analysis – GO terms

GO terms tend to be redundant,
approaches to solve redundancy:

- **DOSE:** disease ontology semantic and enrichment analysis
R package finds enriched disease pathways
- **GoSemSim:** semantic similarity among GO terms and gene products
R package shows how similar are the Gene/Disease ontology terms



Functional analysis – Pathways

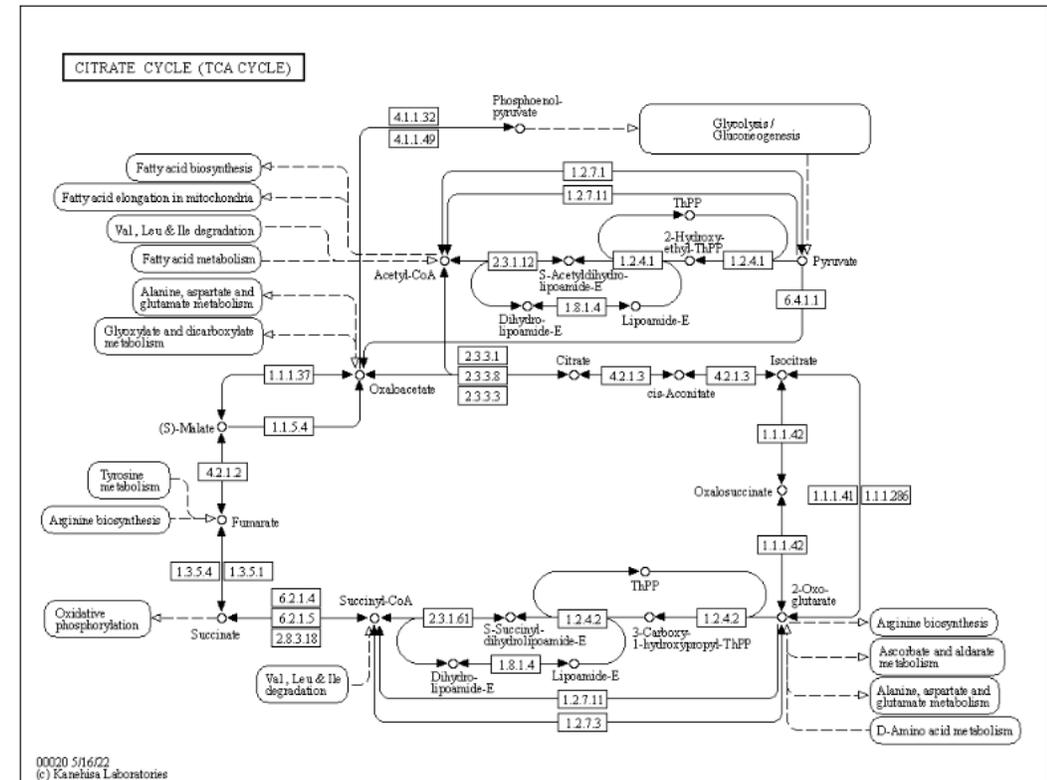
Pathways

Set of genes interacting with each other to perform a specific biological function

KEGG pathway database

- Metabolism
- Genetic information processing
- Environmental Information Processing
- Cellular processes
- Organismal Systems
- Human Diseases

Processing



Exercise

Let's annotate some genes!

- Notebook:
 - *08a_FA_genomic_annotation.Rmd*



How to convert gene IDs

```
## Create background dataset for hypergeometric testing using all genes tested for significance in the results
allCont_genes <- dplyr::filter(res_ids, !is.na(gene)) %>%
  pull(gene) %>%
  as.character()

## Extract significant results
sigCont <- dplyr::filter(res_ids, padj < 0.05 & !is.na(gene))

sigCont_genes <- sigCont %>%
  pull(gene) %>%
  as.character()
```

Now we can perform the GO enrichment analysis and save the results:

```
## Run GO enrichment analysis
ego <- enrichGO(gene = sigCont_genes,
  universe = allCont_genes,
  keyType = "ENSEMBL",
  OrgDb = org.Hs.eg.db,
  ont = "BP",
  pAdjustMethod = "BH",
  qvalueCutoff = 0.05,
  readable = TRUE)
```