

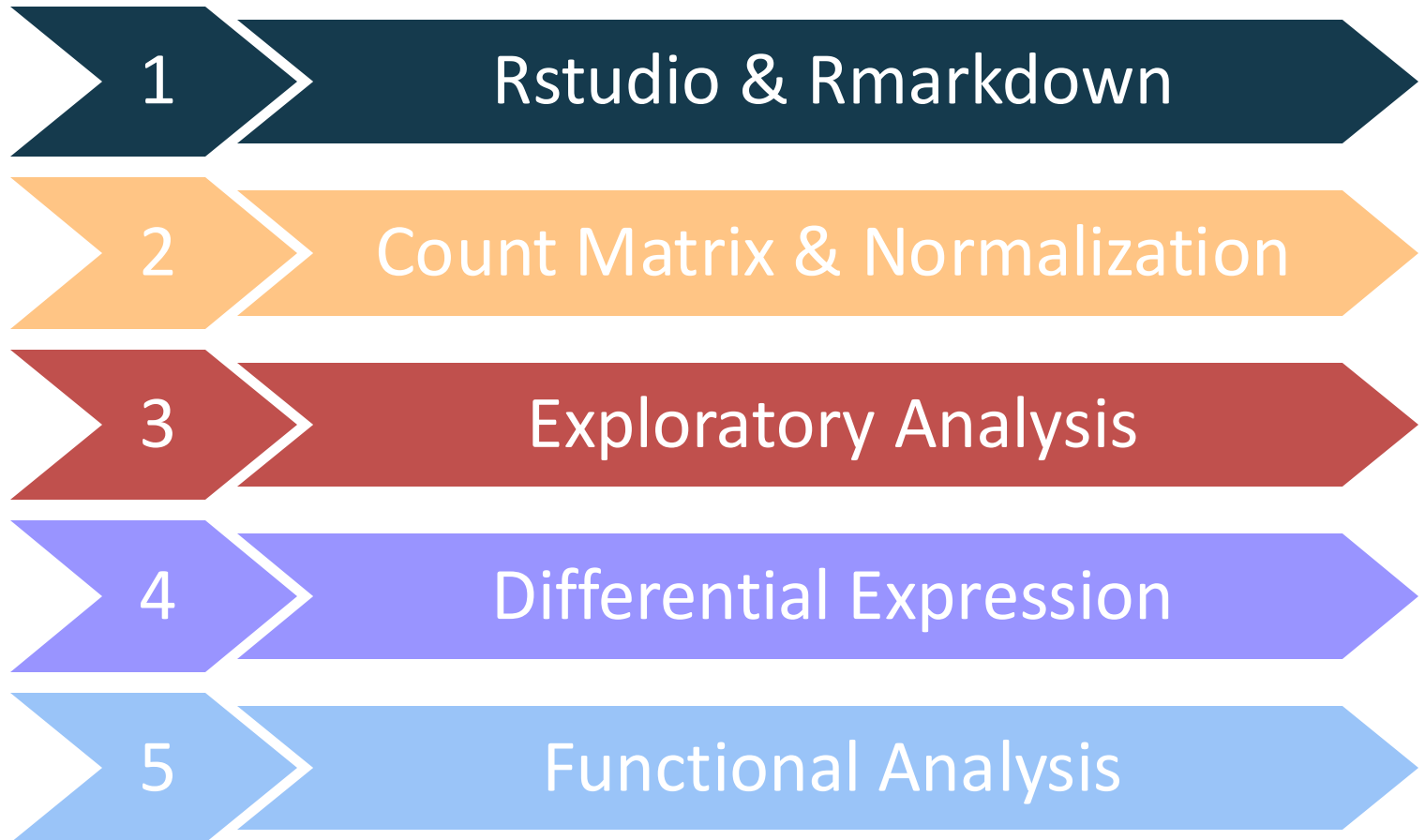
# Functional Analysis

Center for Health Data Science



Health Data Science Sandbox<sup>1</sup>

# Overview



# Functional analysis

- Differential Expression analysis
  - Resulted in **up**- and **down**-regulated genes in each comparison
- Define genes of interest:
  - Treatment vs. Control with  $\text{Log}_2\text{FC} > 1$  and adjusted p-value  $< 0.05$
- What do these genes do?
  - Do they share a **common function**? e.g. Immuno-processes
  - Part of the **same pathway**? e.g. Nucleotide Metabolism

# Functional analysis

1. Differentially  
Expressed Genes

3. Publish!

2. Anything and everything biologically meaningful & interesting:

- Co-expression and interaction
- Gene set enrichment analysis (pathway, GO terms)
- Disease and drug databases
- ...

# Annotation

DESeq2 results:

Gene_ID (Ensembl)	LFC	padj	...
<u>ENSG00000223972</u>	1.101	0.001	...
<u>ENSG00000278267</u>	-4.567	0.045	...

Annotation:

EBI (Ensembl)

HGNC

NCBI

UCSC

Gene_ID (Ensembl)	Gene_name	Entrez_ID	RefSeq_ID	Chr	Start	End	Feature
<u>ENSG00000223972</u>	DDX11L1	NA	NR_046018	1	11869	14409	Protein
<u>ENSG00000278267</u>	MIR6859-1	102466751	NR_106918	1	17369	17436	nc-RNA

+ MANE = Matched Annotation between NCBI and EBI

# Annotation

Genome assembly updated **every few years**

Annotations updated **every few months**

⚠ Genome assembly versions **MUST** match!

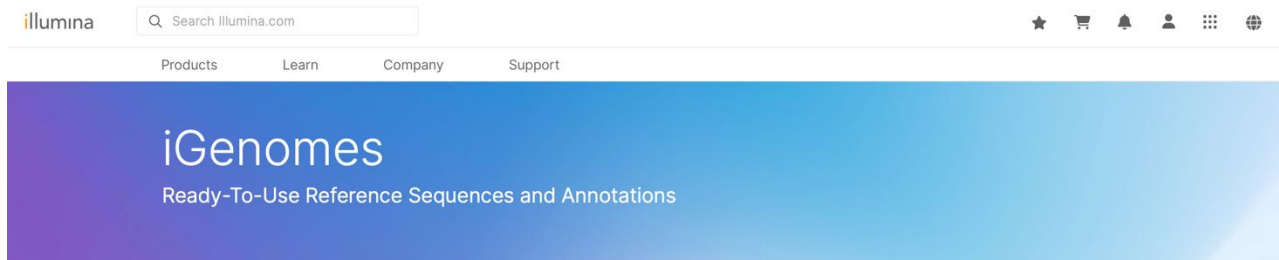
- If you **map to GRCh37**, use **annotations for GRCh37**
- Preferably also use the **same release** for all annotations

Ensemble database annotation releases for GRCh37 build:

```
> listEnsemblArchives()
```

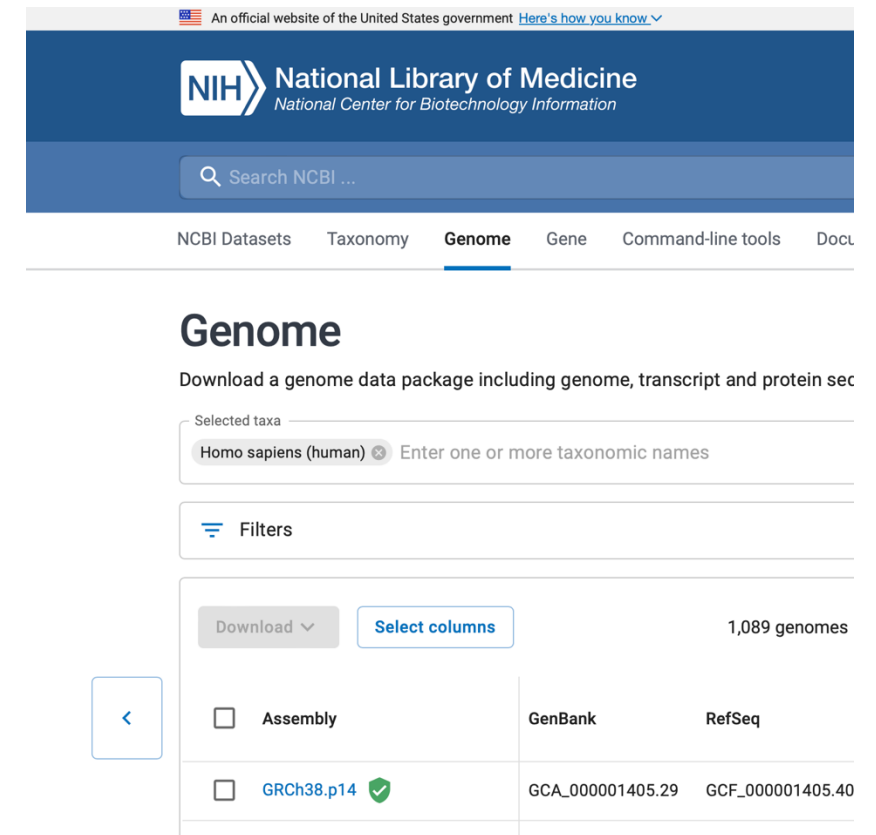
	name	date	url	version	current_release
1	Ensembl GRCh37	Feb 2014	https://grch37.ensembl.org	GRCh37	
2	Ensembl 108	Oct 2022	https://oct2022.archive.ensembl.org	108	*
3	Ensembl 107	Jul 2022	https://jul2022.archive.ensembl.org	107	
4	Ensembl 106	Apr 2022	https://apr2022.archive.ensembl.org	106	
5	Ensembl 105	Dec 2021	https://dec2021.archive.ensembl.org	105	
6	Ensembl 104	May 2021	https://may2021.archive.ensembl.org	104	
7	Ensembl 103	Feb 2021	https://feb2021.archive.ensembl.org	103	
8	Ensembl 102	Nov 2020	https://nov2020.archive.ensembl.org	102	
9	Ensembl 101	Aug 2020	https://aug2020.archive.ensembl.org	101	
10	Ensembl 100	Apr 2020	https://apr2020.archive.ensembl.org	100	
11	Ensembl 99	Jan 2020	https://jan2020.archive.ensembl.org	99	
12	Ensembl 98	Sep 2019	https://sep2019.archive.ensembl.org	98	
13	Ensembl 97	Jul 2019	https://jul2019.archive.ensembl.org	97	
14	Ensembl 96	Apr 2019	https://apr2019.archive.ensembl.org	96	
15	Ensembl 95	Jan 2019	https://jan2019.archive.ensembl.org	95	
16	Ensembl 94	Oct 2018	https://oct2018.archive.ensembl.org	94	
17	Ensembl 93	Jul 2018	https://jul2018.archive.ensembl.org	93	
18	Ensembl 92	Apr 2018	https://apr2018.archive.ensembl.org	92	
19	Ensembl 91	Dec 2017	https://dec2017.archive.ensembl.org	91	
20	Ensembl 80	May 2015	https://may2015.archive.ensembl.org	80	
21	Ensembl 77	Oct 2014	https://oct2014.archive.ensembl.org	77	
22	Ensembl 75	Feb 2014	https://feb2014.archive.ensembl.org	75	
23	Ensembl 54	May 2009	https://may2009.archive.ensembl.org	54	

# Annotation



👉 A resource for downloading matching references + annotations

A more updated resource 👉



# Annotation

## Reference genome = gold standard

Genome assembly GRCh38.p14 reference



- **Chromosome level** assembly
- Contains **chromosomes + scaffolds** with funny names
- Some regions **contain gaps** (telomeres, repetitive regions, centromeres)

## Newer version but less explored

Genome assembly T2T-CHM13v2.0

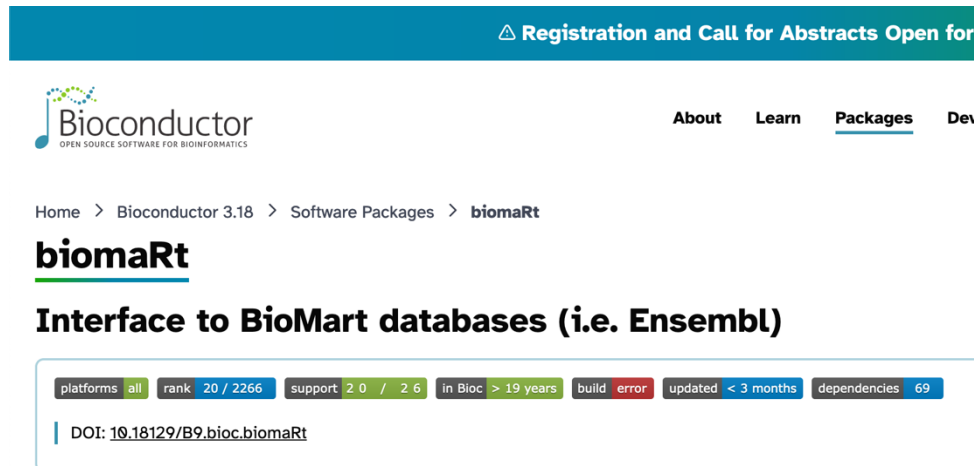


- **Whole genome** assembly
- Contains **only chromosomes**
- **Complete sequence** from telomere to telomere



# Annotation in R

- R packages **biomaRt** or **AnnotationDbi** can be used to convert between identifiers
- You *may* not be able to get the exact release version of a build, but you can likely get one close to it:
  - This could mean some IDs cannot be converted, it is only a few we usually don't care



△ Registration and Call for Abstracts Open for

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About Learn Packages Dev

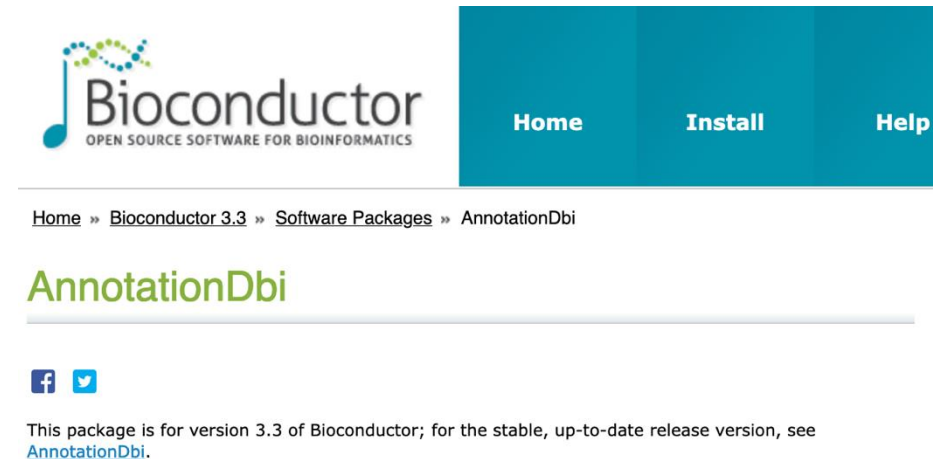
Home > Bioconductor 3.18 > Software Packages > **biomaRt**

**biomaRt**

Interface to BioMart databases (i.e. Ensembl)

platforms all rank 20 / 2266 support 2.0 / 2.6 in Bioc > 19 years build error updated < 3 months dependencies 69

DOI: [10.18129/B9.bioc.biomaRt](https://doi.org/10.18129/B9.bioc.biomaRt)



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Home Install Help

Home » Bioconductor 3.3 » Software Packages » AnnotationDbi

**AnnotationDbi**

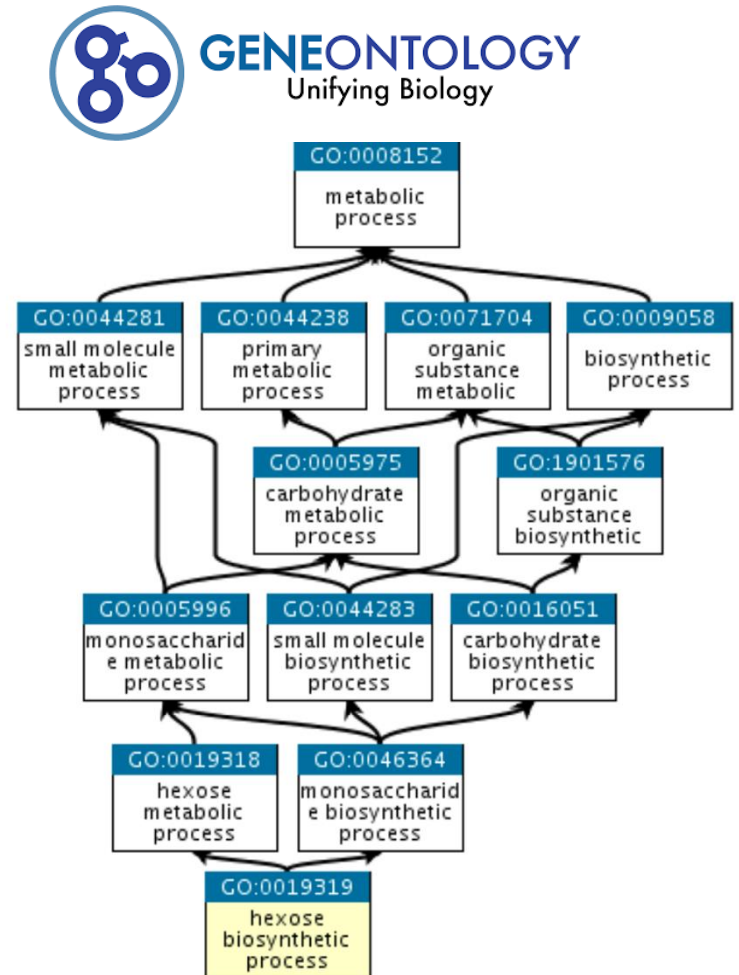
f t

This package is for version 3.3 of Bioconductor; for the stable, up-to-date release version, see [AnnotationDbi](#).

# Functional analysis – GO terms

## Gene Ontology (GO) Term

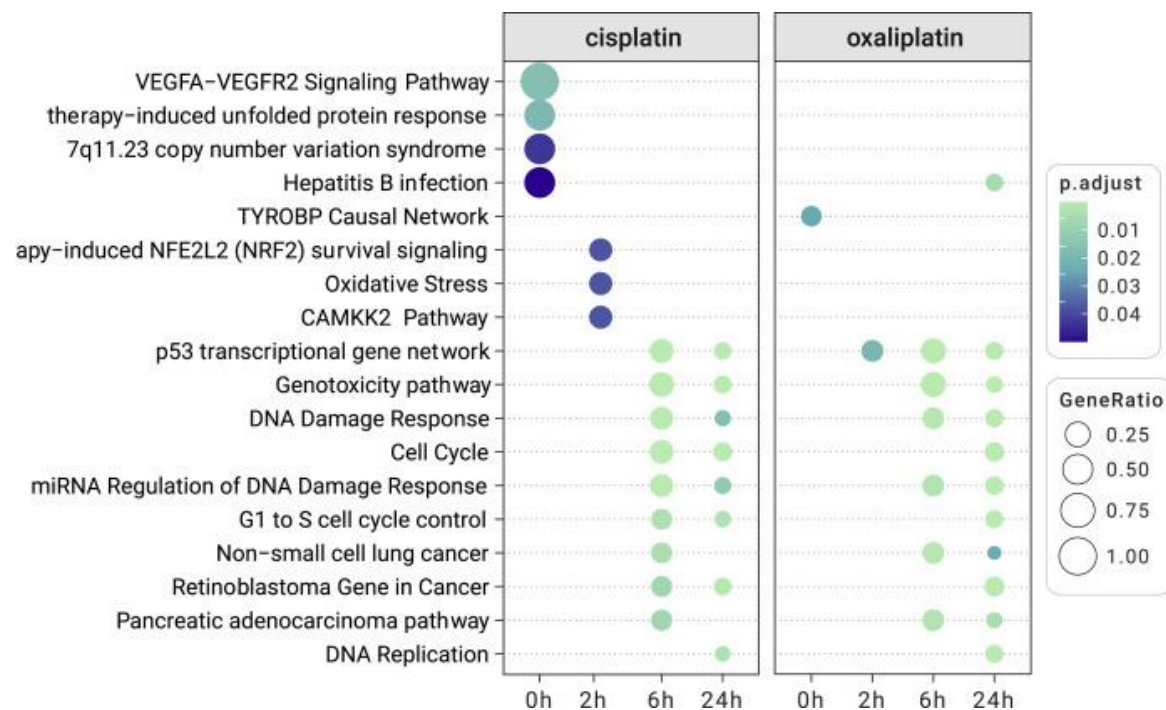
- Formal representation of a body of knowledge in the biological domain
- Genes are annotated to different types of knowledge
  - Biological processes: DNA repair, signal transduction, etc.
  - Molecular function: catalysis, transportation, etc.
  - Cellular component: ribosome, nucleus, etc.



# Functional analysis – GO terms

GO terms tend to be redundant,  
approaches to solve redundancy:

- **DOSE:** disease ontology semantic and enrichment analysis  
R package finds enriched disease pathways
- **GoSemSim:** semantic similarity among GO terms and gene products  
R package shows how similar are the Gene/Disease ontology terms



# Functional analysis – Pathways

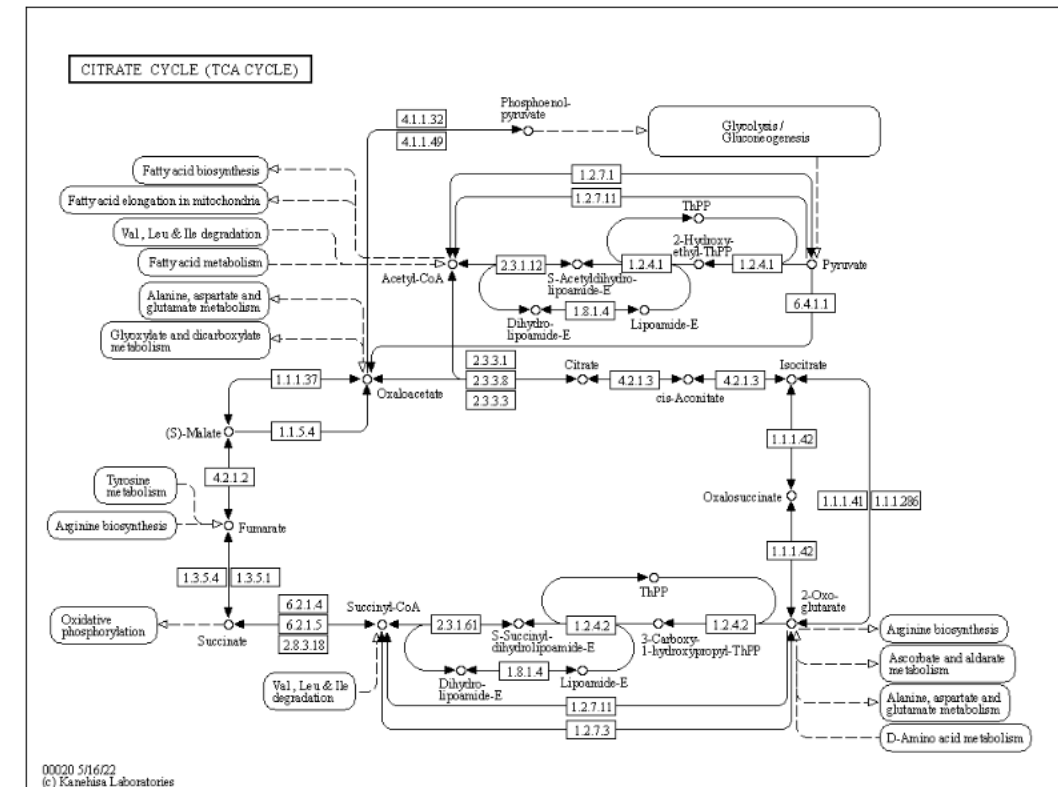
## Pathways

Set of genes interacting with each other to perform a specific biological function

## KEGG pathway database

- Metabolism
- Genetic information processing
- Environmental Information Processing
- Cellular processes
- Organismal Systems
- Human Diseases

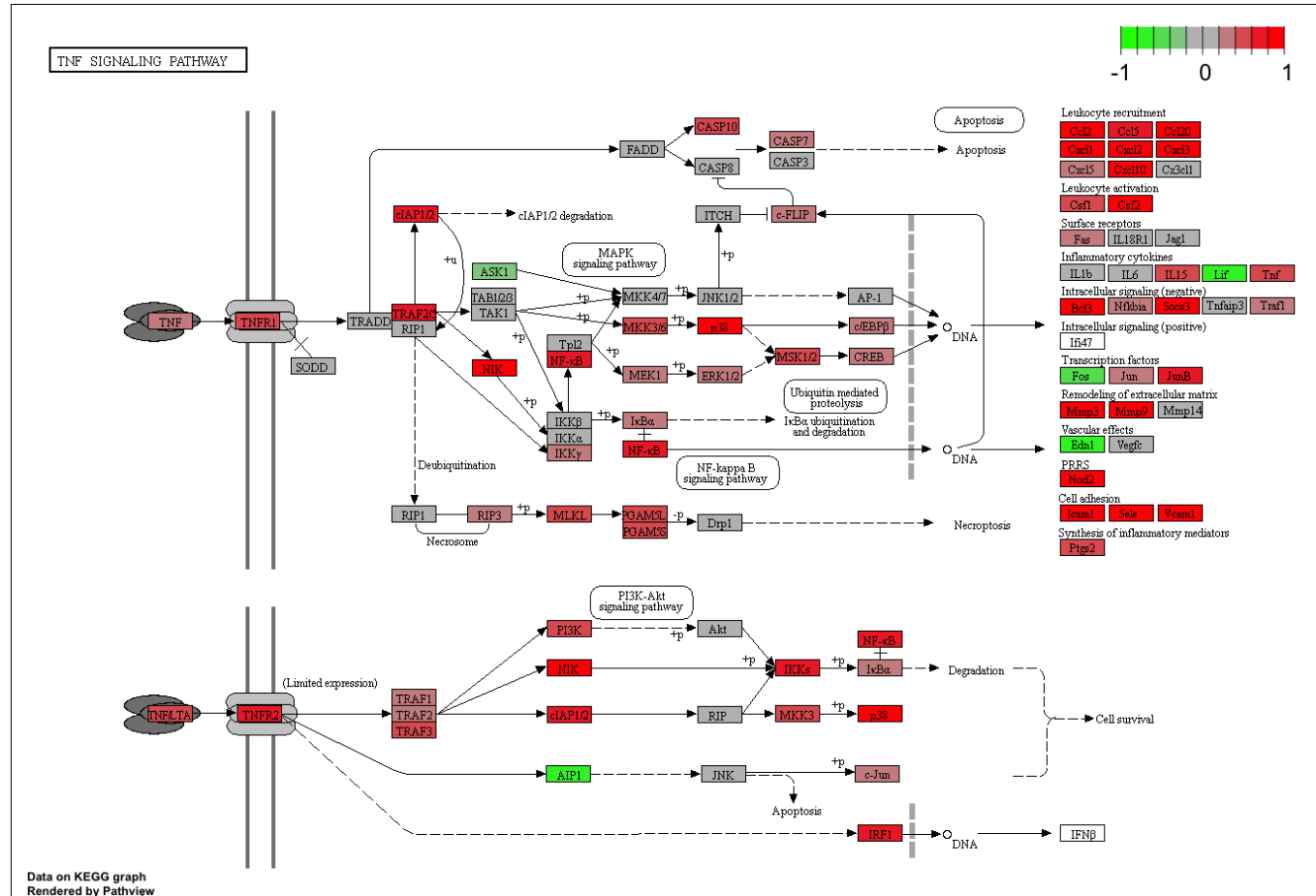
Processing



# Functional analysis – Pathways

## Pathview:

R package that visualizes  
Differentially Expressed (DE)  
genes with their  
log2foldChanges (LFC) within a  
KEGG pathway.



# Functional analysis – Pathways

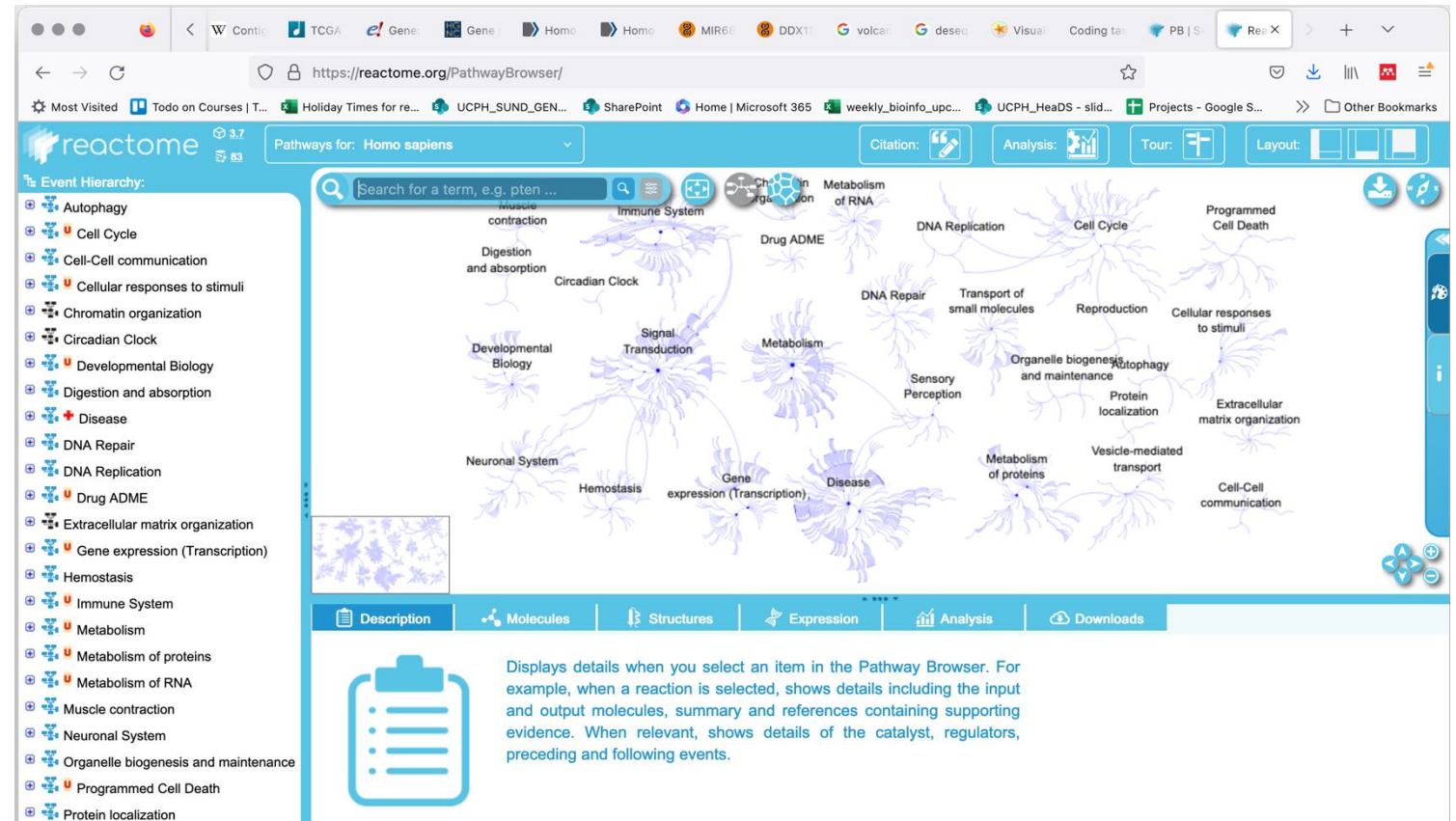
## Reactome:

## Pathway database:

- open-source
- open access
- manually curated
- peer-reviewed

## R-package:

# reactomePA



# Exercise

Let's annotate some genes!

- Notebook:
  - *08a\_FA\_genomic\_annotation.Rmd*



# How to convert gene IDs

```
## Create background dataset for hypergeometric testing using all genes tested for significance in the results
allCont_genes <- dplyr::filter(res_ids, !is.na(gene)) %>%
  pull(gene) %>%
  as.character()

## Extract significant results
sigCont <- dplyr::filter(res_ids, padj < 0.05 & !is.na(gene))

sigCont_genes <- sigCont %>%
  pull(gene) %>%
  as.character()
```

Now we can perform the GO enrichment analysis and save the results:

```
## Run GO enrichment analysis
ego <- enrichGO(gene = sigCont_genes,
  universe = allCont_genes,
  keyType = "ENSEMBL",
  OrgDb = org.Hs.eg.db,
  ont = "BP",
  pAdjustMethod = "BH",
  qvalueCutoff = 0.05,
  readable = TRUE)
```